

Reprint Series
28 September 1984, Volume 225, pp. 1493–1496

SCIENCE

Event-Related Brain Potentials in Boys at Risk for Alcoholism

Henri Begleiter, Bernice Porjesz, Bernard Bihari, and Benjamin Kissin

Event-Related Brain Potentials in Boys at Risk for Alcoholism

Abstract. Recent neurophysiological findings have demonstrated that abstinent chronic alcoholics manifest deficits in event-related brain potentials. To explore possible biological antecedents of alcoholism the present study examined boys at high risk for alcoholism. Event-related brain potentials were recorded from biological sons of alcoholic fathers and matched control boys. Differences in the P3 component of the potentials were obtained between the high-risk and control subjects.

Brain dysfunction or brain damage has been observed with the use of neuropsychological and neuroradiological techniques in chronic alcoholics (1). Studies of evoked brain potentials (EP's) have demonstrated a number of functional aberrations in chronic alcoholics (2). Several investigators have studied auditory brain stem potentials in chronic alcoholics and have reported electrophysiological evidence of increased neural transmission time (3). Moreover, event-related potential (ERP) studies in chronic alcoholics have demonstrated deficits in the P3 component with the use of information processing paradigms (4). The presence of these deficits in the central nervous system has been presumed to reflect the consequence of chronic alcohol abuse (toxic effects of alcohol on the brain, nutritional deficits, or an interaction of alcohol and nutrition-related factors). Although the neurophysiological deficits observed in chronic alcoholics are presumed to be alcohol-related effects, it is possible that some of these deficits may be present in subjects at high risk for alcoholism and therefore antecede the onset of alcohol abuse.

Genetic factors may be involved in the development of alcoholism. Sons of alcoholic fathers represent a special group at high risk for developing alcoholism (5)

even when they are separated from their biological parents soon after birth. Studies of male adoptees indicate that the biological rather than the adoptive parent is predictive of later drinking problems (6). Further evidence for a genetic predisposition comes from twin studies indicating that the concordance rate for alcohol abuse among identical twins is almost double the rate for fraternal twins (7); patterns of alcohol consumption are also highly concordant among identical twins (8). This evidence suggests that a genetic factor may be involved in the presence of neural pathophysiology associated with alcohol abuse.

The identification of a suitable biological marker that is genetically transmitted is important in identifying individuals before the onset of the disease. Moreover, biological markers can provide fundamental data on the etiology of alcoholism. The search for such a marker must focus on a biological variable known to be genetically determined and prevalent in abstinent chronic alcoholics. There is good evidence to indicate that EP waveforms are genetically determined. Monozygotic twins manifest EP waveforms that are as concordant with each other as those obtained from the same individual tested twice (9).

We now report the presence of P3

deficits in the ERP's obtained from subjects at high risk for alcoholism compared with those of control subjects. Twenty-five sons of alcoholic fathers between the ages of 7 and 13 with a mean age of 11.9 (standard deviation, 2.1) were tested. In each case, the father had received the exclusive diagnosis of alcoholism (DSM III) and had at one time or another been in treatment for alcoholism. We excluded boys whose mothers were alcoholics or who had ingested alcohol during pregnancy or drank excessively after giving birth. Only boys without medical problems and without exposure to alcohol or other substances of abuse were included in this study.

The 25 normal control (NC) subjects were boys who were matched for socio-

economic status and age to the high-risk (HR) subjects. The NC group had a mean age of 12.5 years (standard deviation, 2.4) and did not differ significantly in age from the HR group. They were included only if they had no exposure to alcohol or other substances of abuse, and had no history of alcoholism, or other psychiatric disorder in first- or second-degree relatives. Except for alcohol history, the same exclusion criteria were used as for the HR group. All subjects were paid volunteers.

Subjects were seated in a sound-attenuating chamber facing a computer-controlled display (cathode-ray tube). They were told to look at a fixation point displayed in the center of the screen. The experiment consisted of a visual head-

orientation task. The nontarget stimulus was a frequently occurring plain oval presented in the center of the cathode-ray tube to which the subject did not respond ($n = 160$). The target stimulus was an aerial view of the head with the nose and only one ear drawn in, rotated in four different positions: nose up and right ear ($n = 20$), nose up and left ear ($n = 20$), nose down and right ear ($n = 20$), nose down and left ear ($n = 20$). Subjects pressed one of two microswitches as quickly as possible (reaction time) with either the right or left index finger to indicate whether the right or left ear, respectively, was present in the display.

In the "easy" condition, the head was facing forward (nose up on screen), and the left or right ear appeared directly on the side corresponding to the appropriate button. In the "difficult" condition, the head was facing back (nose down on screen), and either the left or right ear appeared on the side of the screen opposite the corresponding button. A total of 240 stimuli were randomly presented—160 nontargets and 80 targets (20 per target condition). The stimuli were 25 msec in duration and subtended 2.9° of arc; interstimulus intervals varied randomly between 2 and 4 seconds. The ERP's and behavioral data were under computer control (10).

Reaction times for easy stimuli were significantly shorter than for difficult stimuli [$F(1, 48) = 110.64, P < 0.0001$]. There were no significant reaction time differences between groups. The number of correct behavioral responses was significantly less for the HR group for easy [$t(48) = 2.76, P < 0.01$] and difficult stimuli [$t(48) = 3.65, P < 0.001$].

All ERP data were subjected to an eye-movement correction procedure (11). ERP's to all stimuli regardless of side of appearance, were combined for easy and difficult targets. Furthermore, ERP's obtained in trials with correct behavioral responses were subjected to a latency corrected average procedure (12) to reduce possible latency jitter. The P3 baseline-to-peak voltage and latency measurements obtained from these latency corrected average data were then analyzed across groups, conditions, and electrodes with a repeated measures analysis of variance (13). No differences in the latency of P3 were obtained between groups. In addition to the baseline-to-peak voltage measurements, the entire raw data set was subjected to a principal component analysis with varimax rotation, through the use of the covariance matrix (PCAV) (13). As the results of our statistical analyses were

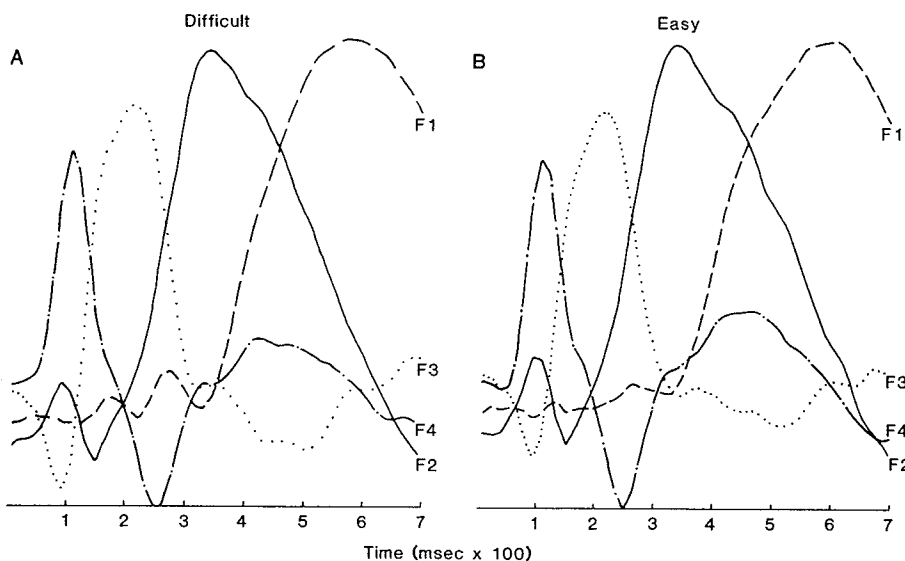


Fig. 1. Factor loadings of the first four factors obtained after principal component analysis with varimax rotation (PCAV) for the difficult (A) and easy (B) target and nontarget stimuli for both groups of subjects (normal control and high risk) at all four electrodes.

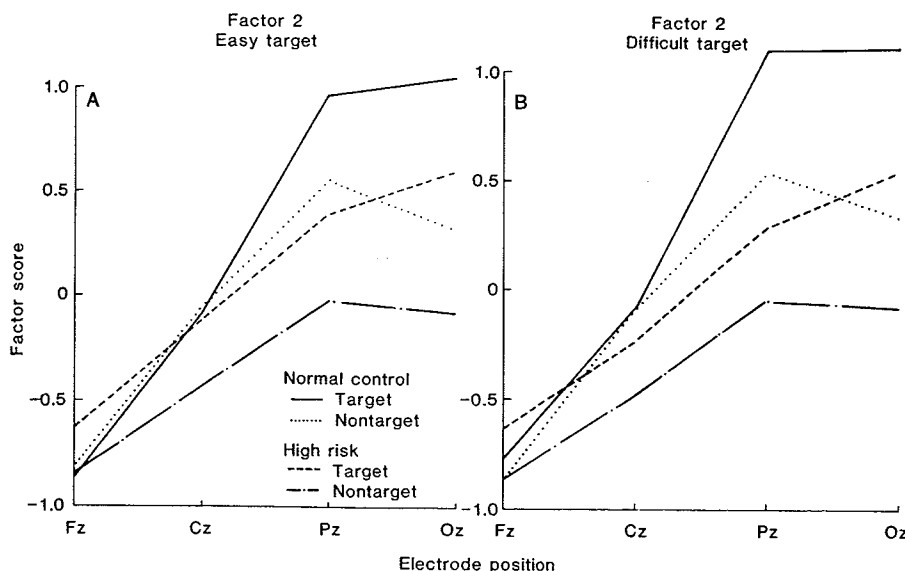


Fig. 2. Factor scores of factor 2 (P3) plotted according to electrode for the easy (A) and difficult (B) target and nontarget PCAV.

identical with both procedures, only the results of the PCAV procedure will be presented here.

Separate PCAV's were performed on the easy target and nontarget ERP's, and the difficult target and nontarget ERP's for the HR and NC groups separately. As the factor structures of the two groups were the same (14), the data from the two groups were combined to perform two separate PCAV's for the easy and difficult conditions. Figure 1 indicates the component loadings of the first four factors obtained for the easy and difficult conditions. The first four factors account for 96.1 percent of the variance for the easy condition and 96.4 percent for the difficult condition. Factor 1 is a rather broad component that peaks at 598.5 msec for the easy and at 570.5 msec for the difficult conditions (slow wave). Factor 2 is maximum at Pz and peaks at 332.5 msec for both the easy and the difficult conditions (P3). The factor scores for each of the four factors were subjected to a repeated measures analysis of variance (13). Factor 1 (slow wave) was significantly different for electrodes [easy: $F(3, 144) = 18.06, P < 0.0001$; difficult: $F(3, 144) = 19.74, P < 0.0001$] and stimuli (target, nontarget) [easy: $F(1, 48) = 43.36, P < 0.0001$; difficult: $F(1, 48) = 53.29, P < 0.0001$], but not for groups. Factor 3 was significantly different for electrodes [easy: $F(3, 144) = 46.95, P < 0.0001$; difficult: $F(3, 144) = 56.67, P < 0.0001$] and stimuli [easy: $F(1, 48) = 21.72, P < 0.0001$; difficult: $F(1, 48) = 19.44, P < 0.0001$], but not for groups. Factor 4 was significantly different only for electrodes [easy: $F(3, 144) = 38.9, P < 0.0001$; difficult: $F(3, 144) = 28.70, P < 0.0001$]. Factor 2 (P3) was also significantly different for electrodes [easy: $F(3, 144) = 57.50, P < 0.0001$; difficult: $F(3, 144) = 64.54, P < 0.0001$] and stimuli [easy: $F(1, 48) = 16.16, P < 0.0002$; difficult: $F(1, 48) = 22.15, P < 0.0001$]. In addition, factor 2 was significantly different between high- and low-risk groups for the difficult condition [$F(1, 48) = 5.22, P < 0.02$].

The component scores for each of the four factors were then subjected to a repeated measures analysis of variance for each electrode separately. Again these results indicated that only factor 2 (P3) differed significantly between groups. This group difference was significant at the parietal lead (where P3 is maximum) for both easy [$F(1, 48) = 6.49, P < 0.01$] and difficult [$F(1, 48) = 9.92, P < 0.002$] conditions, as well as at the occipital lead for the difficult condition [$F(1, 48) = 4.21, P < 0.04$] (Fig. 2).

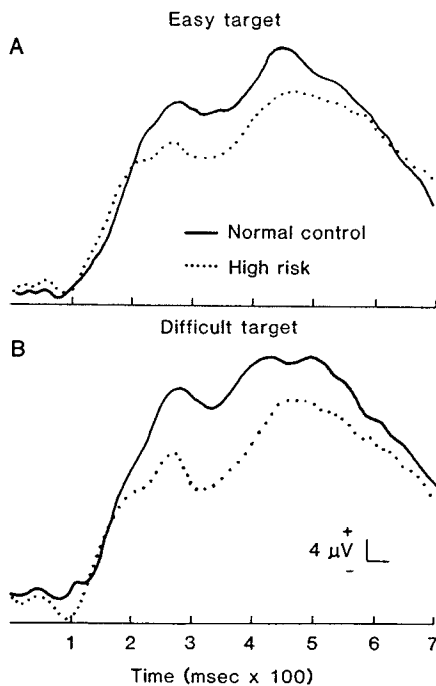


Fig. 3. Grand mean ERP waveforms for the easy (A) and difficult (B) targets at the parietal electrode (Pz).

Figure 3 illustrates the differences between the ERP's obtained from both groups for the easy and difficult targets at the parietal lead, where these differences were greatest.

These findings indicate a significant difference in P3 voltage between boys at high risk for alcoholism and normal control boys, without exposure to alcohol. As reaction times did not differ between groups, it is unlikely that these differences are due to attention deficits. We have obtained similar results in alcoholic patients. Perhaps the differences are due to differences in speed versus accuracy strategies. Although possible, it is unlikely that differences in P3 voltage between these two groups are due to differences in maturational level. Differences in evoked potentials and electroencephalograms have recently been reported between males with some family history of alcoholism and control subjects in response to a challenge dose of alcohol (15). Our findings are particularly striking in that they were obtained without the administration of alcohol.

We have reported similar P3 voltage decrements in abstinent chronic alcoholics with many P3 procedures (2). Recently, we have investigated the reversibility of electrophysiological deficits observed in alcoholics with prolonged abstinence from alcohol. Although we have found improvement in brain stem potential delays, we have not observed any change in P3 voltages (16). These results suggest that rather than being the consequence

of years of heavy drinking, the P3 voltage decrements observed in chronic alcoholics antedate alcohol abuse.

The significance of a reduced P3 component in high-risk boys may be interpreted at two different levels. At the neuroanatomical level, recent intracranial recordings in humans (17) have indicated that the hippocampus and amygdala may make substantial contributions to the P3 potentials recorded from the scalp. In addition, a study using the magnetoencephalographic technique has also suggested the hippocampus as a possible neural generator of this late positive ERP component (18).

At the functional level it is by now well established that the amplitude of P3 indexes stimulus significance (19) and plays a role in memory (20). Thus the significantly reduced P3 amplitude in high-risk boys suggests a reduced capacity to assess significance or allocate the necessary neural resources for encoding a specific event. To the extent that P3 reflects processes involved in revising current representations in working memory, this specific neurophysiological deficit in HR subjects suggests that sons of alcoholics may manifest deficits in memory. This has recently been observed in adolescent sons of alcoholics who performed significantly more poorly than control subjects on memory tasks (21).

The present neurophysiological observations in boys at high risk for alcoholism are striking in that they were obtained without the use of alcohol in sons of alcoholics not previously exposed to alcohol or other drugs of abuse. Our data do not allow us to infer whether the observed P3 deficit in high-risk male children represents a predisposing factor for subsequent alcohol abuse. Longitudinal studies to examine the relationship between the present neurophysiological findings in male children and future patterns of alcohol intake are necessary.

HENRI BEGLEITER
BERNICE PORJESZ
BERNARD BIHARI
BENJAMIN KISSIN

Department of Psychiatry, State
University of New York, Downstate
Medical Center, Brooklyn 11203

References and Notes

1. C. Ryan and N. Butters, in *The Pathogenesis of Alcoholism*, B. Kissin and H. Begleiter, Eds. (Plenum, New York, 1983), pp. 485-538; M. J. Eckardt et al., *Biol. Psychol.* 13, 551 (1978); O. A. Parsons and W. R. Leber, *Alcohol. Clin. Exp. Res.* 5, 326 (1981); L. A. Cala and F. L. Mastaglia, *ibid.*, p. 283; R. E. Tarter, *Int. J. Addict.* 10, 327 (1975); P. L. Carlen et al., *Science* 200, 1076 (1978); A. Wilkinson and P. L. Carlen, *Acta Psychiatr. Scand.* 62, 86 (1980).
2. B. Porjesz and H. Begleiter, *Alcohol. Clin. Exp. Res.* 5, 304 (1981); B. Porjesz and H. Begleiter,

- in *The Pathogenesis of Alcoholism*, B. Kissin and H. Begleiter, Eds. (Plenum, New York, 1983), pp. 415-483.
3. H. Begleiter, B. Porjesz, C. L. Chou, *Science* 211, 1064 (1981); N. S. Chu and K. C. Squires, *Pharmacol. Biochem. Behav.* 13, 241 (1980); H. J. Rosenhamer and B. I. Silfverskiold, *Arch. Neurol.* 37, 293 (1980).
4. B. Porjesz and H. Begleiter, in *Evoked Brain Potentials and Behavior*, H. Begleiter, Ed. (Plenum, New York, 1979), pp. 277-302; ———, R. Garozzo, in *Biological Effects of Alcohol*, H. Begleiter, Ed. (Plenum, New York, 1980), pp. 603-623; H. Begleiter, B. Porjesz, M. Tenner, *Acta Psychiatr. Scand.* 62, 286 (1980); A. Pfefferbaum, T. B. Horvath, W. T. Roth, B. S. Koppell, *Electroencephalogr. Clin. Neurophysiol.* 47, 637 (1979).
5. D. W. Goodwin, *Arch. Gen. Psychiatry* 36, 57 (1979); N. S. Cotton, *J. Stud. Alcohol* 40, 89 (1979); W. M. Grove and R. J. Cadoret, in *The Pathogenesis of Alcoholism*, B. Kissin and H. Begleiter, Eds. (Plenum, New York, 1983), pp. 31-56.
6. D. W. Goodwin *et al.*, *Arch. Gen. Psychiatry* 28, 238 (1973); C. R. Cloniger, M. Bohman, S. Sigurdsson, *ibid.* 38, 861 (1981); R. J. Cadoret and A. Garth, *Br. J. Psychiatry* 132, 252 (1978) (1978).
7. E. Jonsson and T. Nilsson, *Nord. Hyg. Tidskr.* 49, 21 (1968); Z. Hrubec and G. S. Omenn, *Alcohol. Clin. Exp. Res.* 5, 207 (1981).
8. P. Propping, *Hum. Genet.* 35, 309 (1977); C. A. Clifford, D. W. Fuller, H. M. D. Gurling, R. M. Murray, in *Twins Research*, L. Gedda, P. Parisi, W. A. Nance, Eds. (Liss, New York, 1981), pp. 47-52.
9. R. E. Dustman and E. Beck, *Electroencephalogr. Clin. Neurophysiol.* 19, 541 (1965); T. T. Osborne, *Life Sci.* 9, 481 (1970); W. W. Surwillo, *Behav. Genet.* 10, 201 (1980).
10. Monopolar ERP's were recorded with Ag-AgCl electrodes placed on the midline at frontal, central, parietal, and occipital locations (Fz, Cz, Pz, Oz; 10-20 International System). The linked ears served as reference and the nasion served as ground. Eye movements were recorded by electrodes placed above and below the right eye. Trials with excessive eye movements or blinks were rejected before final data analysis. ERP's were sampled by a PDP 11-40 computer for 49 msec before the stimulus (baseline) and for 700 msec (142 points per second; bandwidth 0.01 to 100 Hz) after the stimulus. The digitized data were averaged so that for each individual, each electrode, and each condition (easy or difficult) we derived the ERP's for the target and nontarget stimuli.
11. G. Gratton *et al.*, *Electroencephalogr. Clin. Neurophysiol.* 55, 468 (1983).
12. C. D. McGillem and J. I. Aunon, *IEEE Trans. BioMed. Electron.* 24, 232 (1977).
13. BMDP Statistical Software (Univ. of California Press, Berkeley, 1983).
14. In order to determine if the components derived from the separate PCAV of the two groups represented similar brain activity, the component scores for each extracted component were subjected to stimuli (target or nontarget) by electrode location (Fz, Cz, Pz, Oz) repeated-measures analysis of variance. Because each component manifested similar topographical distribution and relationship to stimulus conditions, electrode locations, and their interactions in both groups, we concluded that each represented similar brain activity.
15. R. Elmasian *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 79, 7900 (1982); V. E. Pollock *et al.*, *Arch. Gen. Psychiatry* 40, 857 (1983).
16. B. Porjesz and H. Begleiter, in *Alcohol and the Brain*, R. Tarter and D. van Thiel, Eds. (Plenum, New York, in press).
17. E. Halgren *et al.*, *Science* 210, 803 (1980).
18. Y. C. Okada, L. Kaufman, S. J. Williamson, *Electroencephalogr. Clin. Neurophysiol.* 55, 417 (1983).
19. E. Donchin, in *Evoked Brain Potentials and Behavior*, H. Begleiter, Ed. (Plenum, New York, 1979), pp. 13-88; W. S. Pritchard, *Psychol. Bull.* 89, 506 (1981); H. Begleiter *et al.*, *Psychophysiology* 20, 95 (1983).
20. T. F. Sanquist, J. W. Rohrbaugh, K. Syndulko, D. B. Lindsley, *Psychophysiology* 17, 568 (1980); E. Donchin, *ibid.* 18, 493 (1981).
21. A. M. Hegedus, A. I. Alterman, R. E. Tarter, *Alcohol. Clin. Exp. Res.*, in press.
22. We thank A. Duggan and B. Skinner for recruiting subjects and D. Chou, J. DeRosa, and M. Gillespie for valuable technical assistance. Supported by PHS grant AA 05524 from the National Institute on Alcohol Abuse and Alcoholism.

28 December 1983; accepted 20 July 1984