

ARTICLE

Involvement of astrocyte metabolic coupling in Tourette syndrome pathogenesis

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Tourette syndrome is a heritable neurodevelopmental disorder whose pathophysiology remains unknown. Recent genome-wide association studies suggest that it is a polygenic disorder influenced by many genes of small effect. We tested whether these genes cluster in cellular function by applying gene-set analysis using expert curated sets of brain-expressed genes in the current largest available Tourette syndrome genome-wide association data set, involving 1285 cases and 4964 controls. The gene sets included specific synaptic, astrocytic, oligodendrocyte and microglial functions. We report association of Tourette syndrome with a set of genes involved in astrocyte function, specifically in astrocyte carbohydrate metabolism. This association is driven primarily by a subset of 33 genes involved in glycolysis and glutamate metabolism through which astrocytes support synaptic function. Our results indicate for the first time that the process of astrocyte-neuron metabolic coupling may be an important contributor to Tourette syndrome pathogenesis.

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INTRODUCTION

Tourette syndrome is a childhood-onset neuropsychiatric disorder characterized by chronic, repetitive involuntary movements and vocalizations, that is, motor and vocal tics. Although genetic factors play an important role in the etiology of Tourette syndrome, and results from twin and family studies have indicated strong familiarity,¹ the underlying pathophysiology is still unclear.² Identifying genetic factors and associated biological mechanisms would be a major step forward, and could provide putative hallmarks for treatment.

To date, only one Tourette syndrome genome-wide association study (GWAS) has been published.³ Their top signal was in the COL27A1 gene with $P=1.85 \times 10^{-6}$, and there were no genetic variants that reached genome-wide significance. In addition, candidate genes from earlier, smaller-scaled candidate gene studies were not replicated, suggesting that these genes are either not causally related to Tourette syndrome or are only important in specific subtypes of Tourette syndrome. A recent study demonstrated that Tourette syndrome is polygenic and likely influenced by hundreds, possibly thousands, of genetic variants with small effects, and that >75% of Tourette syndrome heritability is captured by common genetic variants included in GWAS chips.⁴ An important question that arises from this polygenic nature is whether these thousands of genes of small effect cluster across cellular function or whether they are

distributed randomly across function. Gene-set analysis, which evaluates the combined effect of multiple genetic variants, has been proposed as an efficient method to test functional clustering by identifying sets of functionally related genes underlying polygenic disorders.⁵ In the present study, we applied gene-set analysis for Tourette syndrome using the current largest available Tourette syndrome GWAS data set to elucidate the genetic factors involved in Tourette syndrome. As Tourette syndrome is assumed to be a brain disorder, we restricted ourselves to cellular function related to genes expressed in the brain, and tested sets of genes involved in specific synaptic, astrocytic, oligodendrocyte and microglial functions.

MATERIALS AND METHODS

Subjects and quality control

The gene-set analysis was performed on the raw GWAS genotype data as described in Scharf *et al.*³ Subject inclusion criteria required a Tourette syndrome Classification Study Group diagnosis of definite Tourette syndrome (a DSM-IV-TR diagnosis of Tourette syndrome plus tics observed by an experienced clinician),⁶ and available genomic DNA were extracted either from blood or cell lines. Exclusion criteria consisted of a history of intellectual disability, tardive tourettism or other known genetic, metabolic or acquired tic disorders. European ancestry controls were derived primarily from cohorts of previously genotyped, unselected population controls, as previously described.³

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Principal components computed from the data were used to control for population stratification. After quality control, the full data set contained 1285 cases and 4964 controls, divided into three samples according to genetic ancestry: European ancestry, non-isolates (778 cases, 4414 controls) from North America and Europe; Ashkenazi Jewish (242 cases, 354 controls) from the US and Israel; and French Canadian (265 cases, 196 controls). Quality control was the same as for Scharf *et al*, except with more stringent SNP filters (removing SNPs with: MAF < 0.01 or HWE P -value < $1e-4$ for the European non-isolate sample; MAF < 0.05 or HWE P -value < $1e-3$ for the Ashkenazi/French Canadian isolate samples).

Genotyping and annotation

Genotyping was conducted on the Illumina Human610-Quad v1_B SNP array for the majority of the subjects and on the Illumina HumanCNV370-Duo v1 for 148 cases. Annotation of SNPs to genes was based on NCBI human assembly build 37.3 and dbSNP release 135. SNPs were assigned to genes when they lay between the transcription start and stop sites, with no window around the gene.

Gene-set creation

Because of the neuropsychiatric nature of Tourette syndrome, the gene-set analysis focussed on brain cell-specific gene sets, which were taken from previously published, expert curated gene sets. A total of 96 gene sets containing 4666 different brain-expressed genes were used, divided into four cell-based groups representing synaptic (neuronal), astrocyte, oligodendrocyte and microglia function.

The synaptic gene sets were taken from Ruano.⁷ These were defined on the basis of assignment of subcellular function as determined by previous synaptic protein purification experiments and data mining for synaptic genes and gene function, where genes were considered 'synaptic' on the basis of proteomic analysis of synaptic preparations.^{8–11} This resulted in a subdivision into 17 functional synaptic gene sets, plus one additional gene set of otherwise unassigned synaptic genes.

Glial gene sets (oligodendrocyte, astrocyte and microglial sets) were taken from Goudriaan.¹² Goudriaan *et al* conducted an in-depth literature study to select astrocyte, oligodendrocyte and microglia genes on the basis of microarray gene expression patterns. Specificity was further increased by removing overlap between the three glial cell types, as well as removing general neuronal genes. The resulting lists of cell-specific genes were then subdivided into gene sets using the Gene Ontology biological process annotations, resulting in 30 astrocytic, 29 oligodendrocytic and 19 microglial hierarchically organized gene sets.

Statistical analysis

The gene-set analysis was conducted using JAG.¹³ First, a self-contained test was performed for each gene set, testing for the evidence of association with Tourette syndrome, under the null hypothesis of no association. For gene sets found to be significant after correction for multiple testing, a competitive test was performed to test whether the observed association was stronger than expected by chance for gene sets of the same size. P -values were computed using at least 15 000 permutations for the self-contained tests, and 150 random

matched gene sets (with at least 15 000 permutations each) for the competitive test. In addition, the impact of each gene on the gene-set association was assessed, by computing the change in association when removing that gene from the analysis.

Analyses were performed separately for each of the three ancestry groups described above. The resulting P -values were combined using Stouffer's Z-score method,¹⁴ weighted by the square root of the sample size. Bonferroni correction (and a significance threshold of $\alpha = 0.05$ for corrected P -values) was used within each of the four cell-type-based groups, to compensate for multiple testing.

RESULTS

Gene-set analysis of the synaptic, oligodendrocytic and microglial gene sets uncovered no significant association with Tourette syndrome (Supplementary Tables S1–S3). However, within the astrocyte group, a single gene set, representing the astrocyte carbohydrate metabolism pathway, was found to be significantly associated with Tourette syndrome risk in the self-contained test (corrected $P = 0.04$; Supplementary Table S4). The secondary competitive test was also significant ($P = 0.0067$, based on 150 random matched gene sets).

A follow-up analysis was performed to determine whether the association signal of the astrocyte carbohydrate metabolism gene set might be concentrated within a subset of genes with more specific function. For this purpose, the 85 genes in the gene set were subjected to manual data mining based on published data. This resulted in further specification of this gene set into three specific subprocesses related to (i) astrocyte-neuron metabolic coupling (ANMC; 33 genes, coding for enzymes or transporters involved in glycolysis or glutamine metabolism), (ii) extracellular matrix (EM; 10 genes, coding for ECM proteins or proteins that modify ECM) and (iii) glycosylation (GS; 29 genes, coding for enzymes involved in biosynthesis or degradation of glycoproteins); the 15 remaining genes were combined into a fourth 'miscellaneous' subset (see Supplementary Table S5). Gene-set analysis of these four subsets showed that the association was localized to the 33 genes comprising the ANMC gene set, with a corrected P -value of 0.011 for the self-contained test, and $P = 0.0067$ for the competitive test (Table 1).

We further assessed the effect of each of the 33 genes on the gene-set association (Table 2 and Supplementary Table S6). The results show that none of the individual genes would have survived correction for multiple testing, suggesting that the association of the ANMC gene set is not driven by a single gene but rather is due to the combined effect of multiple genes of similar function.

DISCUSSION

We set out to test the hypothesis that the many genes of small effect thought to underlie Tourette syndrome are clustered across cellular function. Despite the relative modest sample size, our gene-set analysis

Table 1 Results for association with Tourette syndrome from gene-set analyses for four specific subgroups of the astrocyte carbohydrate metabolism gene set

Gene set	No. of genes	No. of SNPs	Corrected self. P	Competitive P
Astrocyte carbohydrate metabolism	85	1200	0.0402	0.0067
Astrocyte-neuron metabolic coupling	33	276	0.0106	0.0067
Extracellular matrix	10	345	0.117	—
Glycosylation	29	385	1	—
Miscellaneous	15	306	1	—

Abbreviations: corrected self. P , P -value from the self-contained test corrected for multiple testing; competitive P , P -value from competitive test. Note that competitive tests were only conducted and interpreted for gene sets that survived multiple testing on the self-contained test.

Table 2 Results for individual genes in astrocyte-neuron metabolic coupling gene set

Gene Symbol	No. of SNPs	Gene P-value	Impact
ME1	26	0.00858	1
ALDH5A1	8	0.00992	0.429
GBE1	20	0.103	0.29
GALM	12	0.0367	0.269
PYGL	7	0.057	0.224
CPS1	29	0.143	0.151
PFKFB3	49	0.196	0.0792
PYGB	4	0.181	0.0605
IDH2	6	0.165	0.0596
ENO1	3	0.196	0.0441
PPP1R1A	3	0.525	0.0305
MDH2	2	0.159	0.0211
CS	1	0.402	0.0198
PYGM	1	0.0659	0.0137
PGM3	3	0.354	0.0014
PHKG1	1	0.497	-0.00595
SLC3A2	3	0.344	-0.00598
PFKFB4	4	0.474	-0.00728
KHK	1	0.506	-0.00737
LDHB	1	0.442	-0.00749
PCK2	2	0.381	-0.00955
SLC2A8	1	0.527	-0.0105
PGM2	12	0.291	-0.0183
GPT	1	0.594	-0.0234
AKR1B1	1	0.312	-0.0296
NANS	3	0.239	-0.0405
PDK4	7	0.486	-0.0542
OGDHL	6	0.606	-0.0691
DHTKD1	5	0.722	-0.0769
PFKM	10	0.478	-0.128
PGM1	15	0.498	-0.156
PC	14	0.62	-0.211
AGL	15	0.589	-0.326

Gene *P*-values are not corrected for multiple testing. The impact reflects the decrease in gene-set significance if that gene is removed from the gene set (positive impact means the gene-set *P*-value increases if the gene is removed, negative impact that the gene-set *P*-value decreases).

revealed a significant association between the astrocyte carbohydrate metabolism pathway and Tourette syndrome. Competitive testing showed that this gene set was more strongly associated to Tourette syndrome than expected for a gene set of that size. This association could be further narrowed down to the ANMC subprocess, and we showed the effect of this gene set was not because of an effect of a single gene, but was because of an overall, combined effect of many genetic variants of small effect. This is the first study to point to the involvement of ANMC function in Tourette syndrome, probably through altered glycogen and glutamate/GABA metabolism, and in line with previously hypothesized mechanisms underlying Tourette syndrome pathogenesis that involve perturbations in the balance between excitatory glutamatergic and inhibitory GABAergic transmission within regulatory cortico-striato-thalamocortical circuits.^{15–17}

The ANMC gene set contains astrocyte-enriched genes involved in various energy metabolism processes that support synaptic function¹⁸ (Figure 1). First, whereas neurons have a low glycolytic rate, astrocytes actively take up glucose from the circulation, store it as glycogen and subsequently convert glycogen to lactate for release into neurons under neuronal command.¹⁸ The ANMC gene set contains *GBE1*,

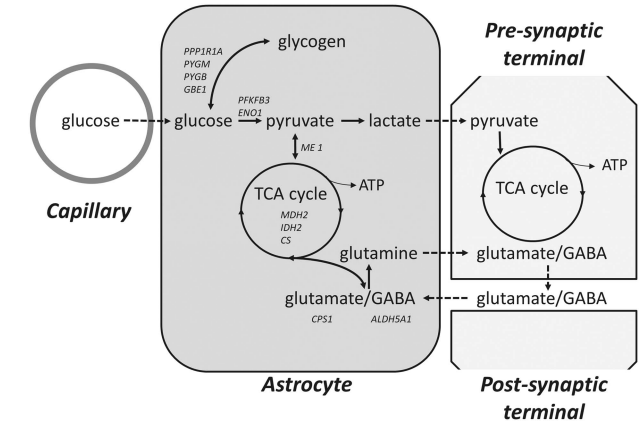


Figure 1 Schematic overview of the astrocyte-neuron metabolic coupling gene set, showing genes positively contributing to the gene-set association with Tourette syndrome. Genetic alterations in astrocyte-neuron metabolic coupling may have downstream effects on various neuronal energy metabolism processes, particularly at synapses: (1) glycolysis-dependent lactate release to the synapse where it is used for ATP generation and (2) glutamate (or GABA) uptake from the synaptic cleft by astrocytes where one part is converted to glutamine and returned to neurons for conversion back to glutamate (or GABA), and another part is used for production of pyruvate and lactate. See main text for further explanation.

PGM3, *PYGM* and *PYGB*, coding for enzymes involved in glycogen storage; *PPP1R1A*, coding for a protein involved in hormonal control of glycogen metabolism; and *PFKFB3* and *ENO1*, coding for glycolytic enzymes for the production of pyruvate and subsequently lactate.

Second, astrocytes take up glutamate (or to a lesser extent GABA) from the synaptic cleft using astrocyte-specific glutamate transporters. A small portion of this glutamate is used in the astrocyte TCA cycle for oxidative energy metabolism and for the production of pyruvate and lactate, in a manner proportional to extracellular glutamate concentration.¹⁹ The larger portion of glutamate is converted to glutamine and shuttled back to neurons for conversion into glutamate (or GABA), independent of extracellular glutamate concentrations and astrocyte energy status.²⁰ The ANMC gene set also contains *CPS1* and *ALDH5A1*, coding for enzymes involved in glutamine and GABA metabolism, respectively; the genes coding for TCA cycle enzymes *MDH2*, *CS* and *IDH2*; and for the key enzyme *ME1*, which links the TCA cycle with the glycolytic pathway. Interestingly, astrocyte glutamate uptake is known to drive glycolysis and subsequent shuttling of lactate to neurons.⁶

Tight regulation of neuronal energy supply by astrocytes in response to synaptic activity is crucial for proper neuronal function.^{18,20} Thus, genetic alterations in glycolysis and glutamate metabolism can have profound influences on astrocyte modulation of synapse function. Such perturbations in the balance between excitatory glutamatergic and inhibitory GABAergic transmission within regulatory cortico-striato-thalamocortical circuits have long been hypothesized as a core defect in Tourette syndrome pathogenesis.^{15–17} Taken together, our findings highlight an often underestimated function of astrocytes in supporting synaptic function and suggest that abnormalities in this process may contribute to the etiology of Tourette syndrome.

CONFLICT OF INTEREST

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TOURETTE SYNDROME ASSOCIATION INTERNATIONAL CONSORTIUM FOR GENETICS

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- O'Rourke JA, Scharf JM, Yu D, Pauls DL: The genetics of Tourette syndrome: a review. *J Psychosom Res* 2009; **67**: 533–545.
- Deng H, Gao K, Jankovic J: The genetics of Tourette syndrome. *Nat Rev Neurol* 2012; **8**: 203–213.
- Scharf JM, Yu D, Mathews CA, Neale BM *et al*: Genome-wide association study of Tourette's syndrome. *Mol Psychiatry* 2013; **18**: 721–728.
- Davis LK, Yu D, Keenan CL *et al*: Partitioning the heritability of Tourette syndrome and obsessive compulsive disorder reveals differences in genetic architecture. *PLoS Genet* 2013; **9**: 1–14.
- Wang L, Jia P, Wolfinger RD, Chen X, Zhao Z: Gene set analysis of genome-wide association studies: methodological issues and perspectives. *Genomics* 2011; **98**: 1–8.
- APA: *Diagnostic and statistical manual of mental disorders*, 4th edn. Washington, DC, USA: American Psychiatric Association, 2000, Text revision (DSM-IV-TR).
- Ruano D, Abecasis GR, Glaser B *et al*: Functional gene group analysis reveals a role of synaptic heterotrimeric G proteins in cognitive ability. *Am J Hum Genet* 2010; **86**: 113–125.
- Li K, Hornshaw MP, van Minnen J, Smalla KH, Gundelfinger ED, Smit AB: Organelle proteomics of rat synaptic proteins: correlation-profiling by isotope-coded affinity tagging in conjunction with liquid chromatography-tandem mass spectrometry to reveal post-synaptic density specific proteins. *J Proteome Res* 2005; **4**: 725–733.
- Li K, Hornshaw MP, Van der Schors RC *et al*: Proteomics analysis of rat brain postsynaptic density: Implications of the diverse protein functional groups for the integration of synaptic physiology. *J Biol Chem* 2004; **279**: 987–1002.
- Fernández E, Collins MO, Uren RT *et al*: Targeted tandem affinity purification of PSD-95 recovers core postsynaptic complexes and schizophrenia susceptibility proteins. *Mol Syst Biol* 2009; **5**: 269.
- Emes RD, Pocklington AJ, Anderson CN *et al*: Evolutionary expansion and anatomical specialization of synapse proteome complexity. *Nat Neurosci* 2008; **11**: 799–806.
- Goudriaan A, de Leeuw C, Ripke S *et al*: Specific glial functions contribute to schizophrenia susceptibility. *Schizophrenia Bull* 2014; **40**: 925–935.
- Lips ES, Cornelisse LN, Toonen RF *et al*: Functional gene group analysis identifies synaptic gene groups as risk factor for schizophrenia. *Mol Psychiatry* 2012; **17**: 996–1006.
- Whitlock MC: Combining probability from independent tests: the weighted Z-method is superior to Fisher's approach. *J Evol Biol* 2005; **18**: 1368–1373.
- Albin RL, Mink WM: Recent advances in Tourette syndrome research. *Trends Neurosci* 2006; **29**: 175–182.
- Singer HS, Morris C, Grados M: Glutamatergic modulatory therapy for Tourette syndrome. *Med Hypotheses* 2010; **74**: 862–867.
- Adamczyk A, Gause CD, Sattler R *et al*: Genetic and functional studies of a missense variant in a glutamate transporter, SLC1A3, in Tourette syndrome. *Psychiatr Genet* 2011; **21**: 90–97.
- Bélanger M, Allaman I, Magistretti PJ: Brain energy metabolism: focus on astrocyte-neuron metabolic cooperation. *Cell Metab* 2011; **14**: 724–738.
- Schousboe A, Bak LK, Madsen KK, Waagepetersen HS: *Amino acid neurotransmitter synthesis and removal*; in Kettenman H, Ransom BR (eds). New York, NY, USA: Neuroglia, 2013, pp 443–456.
- Barros LF: Metabolic signaling by lactate in the brain. *Trends Neurosci* 2013; **36**: 396–404.

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